

Online ISSN: 2682-2628
Print ISSN: 2682-261X

IJC CBR

INTERNATIONAL JOURNAL OF
CANCER AND BIOMEDICAL RESEARCH

<https://jcbr.journals.ekb.eg>

Editor-in-chief

Prof. Mohamed Labib Salem, PhD

**The role of the hepatocyte nuclear factor 4
alpha HNF4 α and the homeobox protein CDX2
immunomarkers in diagnosis of metastatic
colorectal carcinoma**

Alaa I. Amer, Basma S. Amer



PUBLISHED BY
EACR EGYPTIAN ASSOCIATION
FOR CANCER RESEARCH
Since 2014

**International Journal of Cancer & Biomedical Research
(IJCBR) <https://jcbr.journals.ekb.eg>**

IJCBR is an Int. journal published by the Egyptian Society of Cancer Research (EACR, established in 2014, <http://eacr.tanta.edu.eg>) and sponsored by the Egyptian Knowledge Bank (EKB: www.ekb.eg).

IJCBR has been approved by the Supreme Council of Universities, Egypt with score 7 (<http://egjournal.scu.eg>). The journal is cited by google scholar and registered by Publons (<https://publons.com>). The journal has recently been evaluated in 2020 by Nature Springer with a good standing.

Scope of IJCBR

- Drug discovery from natural and synthetic resources
- BioMedical applications of nanotechnology
- Sem cell biology and its application
- Basic and applied biotechnology
- Inflammation and autoimmune diseases
- In silico models and bioinformatics
- In vitro and In vivo preclinical animal models
- Cellular and molecular cancer biology
- Cancer Immunology and Immunotherapy
- New methods for prediction, early detection, diagnosis prognosis and treatment of diseases.
- Immunology in health and diseases
- Anti-microbial defense mechanisms
- Cellular and molecular physiology and pathology of diseases

IJCBR Editor,
Prof. Mohamed Labib Salem, PhD
Professor of Immunology
Faculty of Science, Tanta University, Egypt

The role of the hepatocyte nuclear factor 4 alpha HNF4 α and the homeobox protein CDX2 immunomarkers in diagnosis of metastatic colorectal carcinoma

Alaa I. Amer, Basma S. Amer

Department of Pathology, Faculty of Medicine, Tanta University, Egypt

ABSTRACT

Background: Detection of the primary site of metastatic carcinoma of unknown origin is necessary to help in the choice of the treatment. CDX2 is routinely used in metastatic adenocarcinomas cases for identifying the gastrointestinal origin. HNF4 α is a new immunohistochemical marker with a few studies showing that it is expressed in a majority of colorectal adenocarcinomas. **Aim:** To determine the expression of HNF4 α and CDX2 in metastatic carcinoma cases for detection of gastrointestinal origin. **Material and methods:** We assessed HNF4 α and CDX2 in 60 metastatic carcinoma cases in different organs, which were diagnosed retrospectively. HNF4- α and CDX-2 expressions were evaluated by light microscopic examination of all tissue sections by two different pathologists. The cut-off for positivity of HNF4 α and CDX-2 was 1% of stained nucleus of metastatic tumor cells. The sensitivity, specificity, negative predictive value (NPV), positive predictive value (PPV) and accuracy of two markers and P value were calculated. **Results:** The calculated sensitivity, specificity, positive predictive value, negative predictive value and accuracy of HNF4 α were 90%, 82.5%, 72%, 94.3% and 85% respectively for diagnosis of colorectal carcinoma metastasis from other metastatic carcinomas. The sensitivity, specificity, positive predictive value, negative predictive value and accuracy of CDX2 were 80%, 72.5%, 59.3%, 87.9% and 75% respectively. The P-value for comparing between their expression in metastatic adenocarcinoma from lower GIT and each other groups is less than 0.05. **Conclusions:** We concluded that HNF4 α can be used as a supplementary marker with CDX2 to identify metastatic colorectal adenocarcinoma. HNF4 α expression is a highly specific and sensitive marker of colorectal origin.

Keywords: CDX2, HNF4 α , NPV, PPV

Editor-in-Chief: Prof. M.L. Salem, PhD - Article DOI: 10.21608/IJCBR.2020.33334.1048

ARTICLE INFO



Article history

Received: June 20, 2020

Revised: July 17, 2020

Accepted: July 27, 2020

Correspondence to:

Basma S. Amer
Department of Pathology,
Faculty of Medicine,
Tanta University, Egypt
Mobile: 01016660737
Email:
basmasaedamer@yahoo.com

INTRODUCTION

Metastatic carcinoma from an unknown primary site is a popular clinical problem, so the identification of primary tumor site needs clinical examination and further investigations (Varadhachary, 2007). The choice of treatment is depending on detection of the primary site of metastatic carcinoma of unknown primary, as this help in several steps in the identifying of the primary site. Clinical examination, including age, sex, past and family history with the site of metastases can give an initial indication. The histopathological results are beneficial, but cannot distinguish between several primary

carcinomas. Immunohistochemistry is the most useful tool in the diagnosis of metastatic carcinoma of unknown primary origin (Park et al., 2007).

CDX2 is a homeobox gene that plays a role in the differentiation of intestinal epithelial cells. These genes have an important function in the monitoring of embryonal development of the alimentary tract. CDX2 is participated in the cell proliferation, differentiation, and apoptosis of the intestine. CDX2 is a transcription factor, which high expression of multiple gene products linked with maturation of epithelial cells of the intestine. (Li and Folpe, 2004).

Hepatocyte nuclear factor 4-alpha (HNF4- α) is a nuclear transcription factor with essential roles in the development, differentiation, and metabolism of liver and gastrointestinal tract (Babeu and Boudreau, 2014). The organization of HNF4- α expression and activity is a complicated process. HNF4- α regulation comes at multiple levels: Epigenetic, transcriptional and post-translational. Immunohistochemical detection of HNF4- α has a valuable diagnostic and prognostic role (Lu, 2016). HNF4- α is expressed in the hepatic, gastrointestinal, pancreatic and renal tissues. No immunostaining is noted in the thyroid, lung, breast, ovarian, endometrial and prostatic tumors (Tanaka et al., 2006).

HNF4- α has a significant function in the development of the colon. It is involved in the differentiation, enteric metabolism, and epithelial intestinal adhesion (Garrison et al., 2006) In the gut, HNF4- α plays a crucial role in the regulation of intestinal genes. Inflammatory bowel disease (IBD) and carcinoma of the colon showed alternation of HNF4- α expression (Chellappa et al., 2016).

HNF4 belongs to the superfamily of nuclear receptors, one of the largest families of transcription factors. In mammals, two genes encode the different HNF4a (NR2A) and HNF4g (NR2A2) isoforms. HNF4a is expressed in the liver, stomach, pancreas, kidney and intestine, 14,15 whereas HNF4g is mostly intestinal.

In vivo and in vitro studies have shown that HNF4a has pleiotropic roles in the liver. 17 In the gut, HNF4a has important functions and is a key regulator of enterocytic markers. 18,19 Conditional knockout of the Hnf4a gene in the mouse embryonic colon causes lethality because of the perturbation of organogenesis, cytodifferentiation and gene expression, 20 and its ablation in the adult intestine reveals its involvement in homeostasis, cell architecture and barrier function HNF4 belongs to the superfamily of nuclear receptors, one of the largest families of transcription factors. In mammals, two genes encode the different HNF4a (NR2A) and HNF4g (NR2A2) isoforms. HNF4a is expressed in the liver, stomach, pancreas, kidney and intestine, 14,15 whereas HNF4g is mostly intestinal. 16 In vivo and in vitro

studies have shown that HNF4a has pleiotropic roles in the liver. 17 In the gut, HNF4a has important functions and is a key regulator of enterocytic markers. 18,19 Conditional knockout of the Hnf4a gene in the mouse embryonic colon causes lethality because of the perturbation of organogenesis, cytodifferentiation and gene expression, 20 and its ablation in the adult intestine reveals its involvement in homeostasis, cell architecture and barrier function HNF4 belongs to the superfamily of nuclear receptors, one of the largest families of transcription factors. In mammals, two genes encode the different HNF4a (NR2A) and HNF4g (NR2A2) isoforms. HNF4a is expressed in the liver, stomach, pancreas, kidney and intestine, 14,15 whereas HNF4g is mostly intestinal. 16 In vivo and in vitro studies have shown that HNF4a has pleiotropic roles in the liver. 17 In the gut, HNF4a has important functions and is a key regulator of enterocytic markers. 18,19 Conditional knockout of the Hnf4a gene in the mouse embryonic colon causes lethality because of the perturbation of organogenesis, cytodifferentiation and gene expression, 20 and its ablation in the adult intestine reveals its involvement in homeostasis, cell architecture and barrier function

Detection of the primary site of metastatic carcinoma of unknown origin is important for proper assessment and management of the metastatic carcinoma patients. So this study aims to determine the expression of HNF4- α and CDX2 in metastatic carcinoma from different primary sites, with the evaluation of sensitivity and specificity of these markers.

MATERIAL AND METHOD

Patients and specimens: Metastatic carcinoma tissues in different organs (breast, thyroid, breast and lymph node). All specimens used in the study were obtained from Pathology Department, Tanta University and private laboratories. None of the patients received preoperative treatment, either radiotherapy or chemotherapy. Metastatic carcinoma tissues were obtained from 60 patients.

IHC analysis: Formalin-fixed paraffin-embedded tissue sections (4- μ m) were put on to APES-coated glass slides (Chenglin, Shanghai,

China). Slides were dewaxed in xylene twice for 10 min and rehydrated through graded ethanol. Antigen retrieval was performed in 0.01 mol/l citrate buffer (pH 6.0) by boiling for 10 min. Endogenous peroxidase activity was suppressed with 3% hydrogen peroxide for 10 min. After washing with phosphate-buffered saline (PBS), the slides were blocked with 5% BSA for 30 min at 37°C. Sections were incubated with primary rabbit monoclonal antibody to human CDX-2 ((RM-2116-S0) (Labvision)) and primary rabbit monoclonal antibody to human HNF4-a ((p 41235) (cell signaling technology)) at 4°C overnight, in a humidified chamber. After washing three times with PBS, sections were incubated for 30 min with the secondary antibody (peroxidase goat anti-mouse IgG; dilution, 1:300; catalogue no. 32230; Zymed, San Diego, CA, USA). After washing for three times in PBS, 3,3'-diaminobenzidine (as chromogen) was used. Slides were counter-stained with hematoxylin for 1 min. Sections were not incubated with the primary antibody, used as negative controls.

Immunohistochemical evaluation: HNF4- α and CDX-2 expressions were evaluated by light microscopic examination of all tissue sections by two different pathologists. The cut-off for positivity of HNF4-a and CDX-2 was 1% of stained nucleus of colorectal adenocarcinoma cells. So HNF4- α and CDX-2 Immunoreactivity was scored as negative (0, no immunostaining) or positive (Dabir et al., 2018; Saandi et al., 2013).

Statistical analysis: Statistical presentation and analysis of present study were conducted, using the mean, standard deviation, and chi-square test, person correlation by SPSS (version 15) software.

RESULTS

HNF4- α Expression (Figures 2,6,10 & Tables 1,2)

HNF4- α expression was identified in the nucleus of metastatic tumor cells of lower and upper gastrointestinal tract carcinoma and it was identified in the nucleus of metastatic non adenocarcinoma (from squamous cell carcinoma), and the positivity of tumor cells was only evaluated. It was expressed in metastatic tumor cells of 18 out of 20 cases metastatic adenocarcinoma from the colon (90%). Four

cases of metastatic adenocarcinoma from upper gastrointestinal tract showed positive expression (20%). Two out of 10 cases of metastatic adenocarcinoma not come from gastrointestinal tract showed positive expression (20%). One case out of 10 cases of squamous cell carcinoma showed positive immunostaining for HNF4- α (10%).

CDX-2 Expression (Figures 3,5 and Tables 1,2)

CDX-2 expression was identified in the nucleus of metastatic tumor cells of lower and upper gastrointestinal tract carcinoma and it was identified in the nucleus of metastatic non adenocarcinoma (from squamous cell carcinoma), and the positivity of tumor cells was only evaluated. It was expressed in metastatic tumor cells of 16 cases out of 20 cases metastatic adenocarcinoma from the colon (80%). Five cases of metastatic adenocarcinoma from upper gastrointestinal tract showed positive expression (25%). Four cases out of 10 of metastatic adenocarcinoma not come from gastrointestinal tract showed positive expression (40%). Two cases out of 10 cases of squamous cell carcinoma showed positive immunostaining for CDX-2 (20%).

The sensitivity, specificity, negative predictive value (NPV), positive predictive value (PPV), accuracy of two markers and p value for comparing between their expression in metastatic adenocarcinoma from lower GIT and each other group are shown in Table 2. The positive expression of HNF4- α showed 90% sensitivity however, specificity was 82.5%. The positive expression of CDX-2 showed 80% sensitivity and 72.5% specificity. So HNF4- α and CDX-2 positive expression can be used to diagnose metastatic colorectal carcinoma in different tissue.

DISCUSSION

Recently immunomarkers have been examined for their diagnostic role. Results remain inconclusive and controversial, and up till now, no molecular markers can be used in the routine assessment of metastatic carcinoma of unknown origin. The difficulty of the diagnosis of metastatic carcinoma of the unknown is the detection of primary site. This is a challenge to oncologists and pathologists, the detection of

Table 1. CDX-2 and HNF4-a expression in relation to different types of metastatic adenocarcinoma and metastatic nonadenocarcinoma.

Type	Number	Age	Sex		CDX-4		HNF4-a	
			Male	Female	+	-	+	-
Metastatic adenocarcinoma from lower GIT	20	30	15	5	16	4	18	2
Metastatic adenocarcinoma from upper GIT	20	50	15	5	5	15	4	16
Metastatic adenocarcinoma from non GIT source	10	55	7	3	4	6	2	8
Metastatic non adenocarcinoma	10	78	7	3	2	8	1	9

Table 2. Sensitivity, specificity, PPV, NPV, accuracy, and p-value of HNF4-α and CDX-2 to diagnose metastatic colorectal carcinoma in different tissue

	Sensitivity (%)	Specificity (%)	PPV (%)	NPV (%)	Accuracy
HNF4-α(+ve)	90%	82.5%	72%	94.3%	85%
CDX-2 (+ve)	80%	72.5%	59.3%	87.9%	75%

	Metastatic adenocarcinoma from lower GIT (n= 20)		Metastatic adenocarcinoma from upper GIT (n= 20)		Metastatic adenocarcinoma from non GIT source (n= 10)		Metastatic non adenocarcinoma (n= 10)	
	No.	%	No.	%	No.	%	No.	%
CDX-4								
+	16	80.0	5	25.0	4	40.0	2	20.0
-	4	20.0	15	75.0	6	60.0	8	80.0
p			<0.001*		0.045*		0.004*	
HNF4-α								
+	18	90.0	4	20.0	2	20.0	1	10.0
-	2	10.0	16	80.0	8	80.0	9	90.0
p			<0.001*		<0.001*		<0.001*	

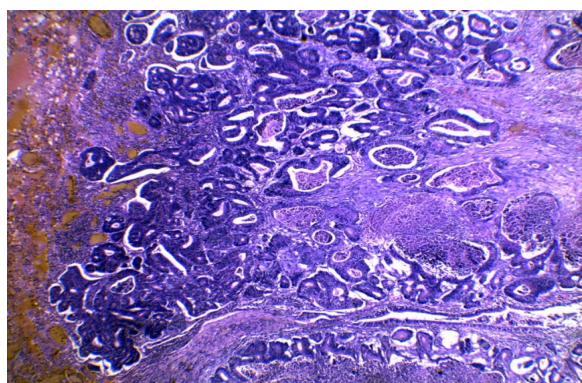


Figure 1. Metastatic grade (II) carcinoma from upper gastrointestinal tract carcinoma case in thyroid tissue (x40).

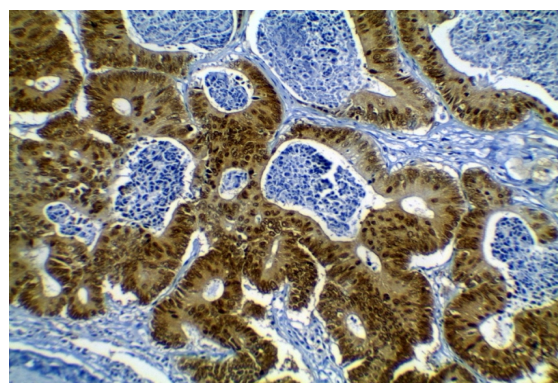


Figure 3. Metastatic grade (II) carcinoma from upper gastrointestinal tract carcinoma case in thyroid tissue showing positive CDX-2 nuclear immunostaining (x200).

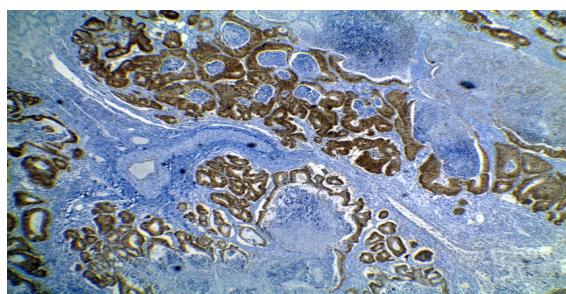


Figure 2. Metastatic grade (II) carcinoma from upper gastrointestinal tract carcinoma case in thyroid tissue showing positive HNF4-α nuclear immunostaining (x40).

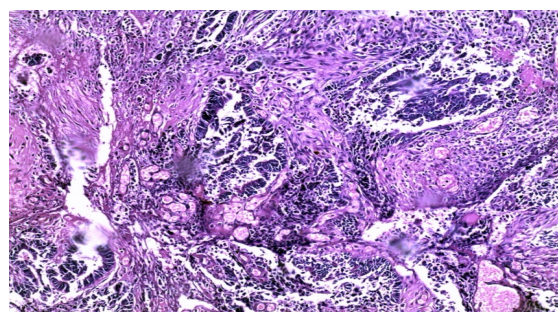


Figure 4. Metastatic grade (II) carcinoma from colon carcinoma case in breast tissue (x100).

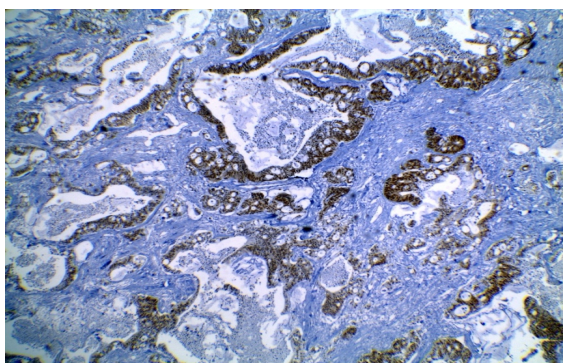


Figure 5. Metastatic grade (II) carcinoma from colon carcinoma case in breast tissue showing positive CDX-2 nuclear immunostaining (x40).

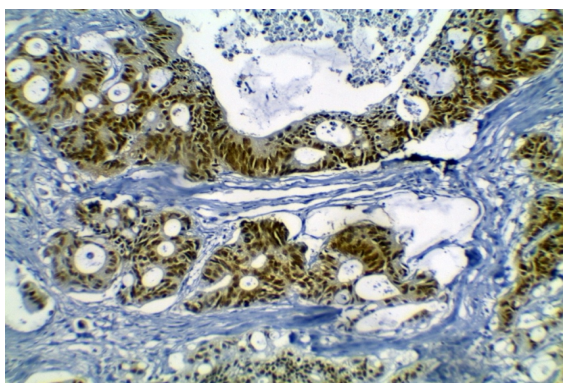


Figure 6. Metastatic grade (II) carcinoma from colon carcinoma case in breast tissue showing positive HNF4-α nuclear immunostaining (x200).

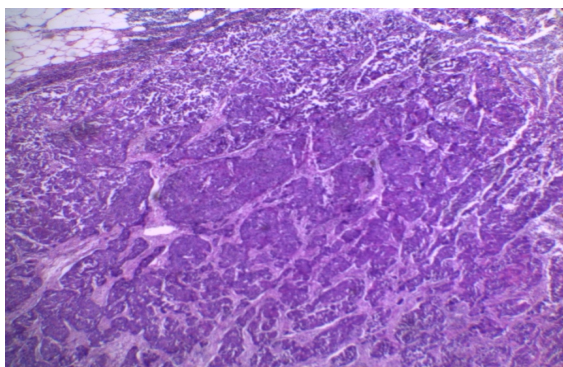


Figure 7. Metastatic grade (II) carcinoma from squamous cell carcinoma case in lymph node tissue (x40).

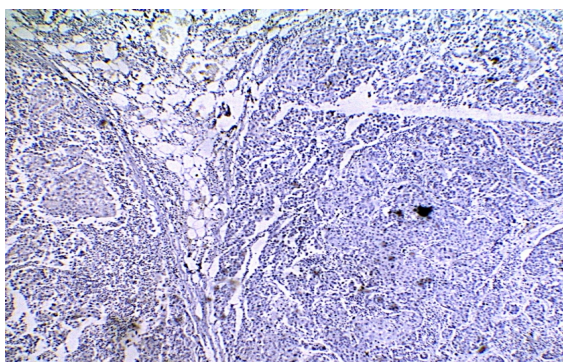


Figure 8. Metastatic grade (II) carcinoma from squamous cell carcinoma case in lymph node tissue (x40).

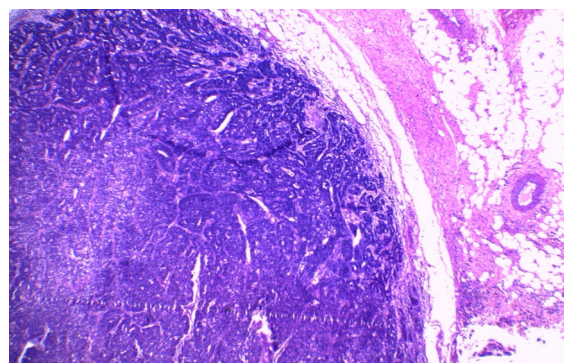


Figure 9. Metastatic grade (II) carcinoma from colon carcinoma case in lymph node tissue (x40).

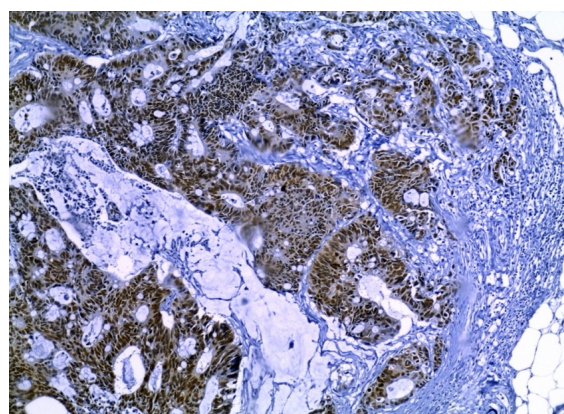


Figure 10. Metastatic grade (II) carcinoma from colon carcinoma case in lymph node tissue showing positive HNF4-4 nuclear immunostaining (x100).

the primary site has important diagnostic and therapeutic outcomes. The clinical, pathological correlation and a panel of immunostains are useful to determinate the origin for proper management of the patients (Park et al., 2007). Human CDX2 protein is one of homeobox genes that encodes an intestinal specific transcription factor. CDX2 protein can regulate the development of the intestine and is showed nuclear expression of epithelial intestinal cells through the intestinal tract in embryonal and post-natal life. The expression of CDX2 is highly restricted to intestinal epithelium (Silberg et al., 2000). Recently, the sensitivity and specificity of CDX2 protein as immunomarker of metastatic colonic carcinoma is evaluated in many types of research. CDX2 is a useful marker and can be included in the IHC panel, as it is a relatively sensitive and specific marker for colorectal adenocarcinomas (Borrisholt et al., 2013).

In the current study, CDX2 was expressed in 16 of 20 (80%) colorectal adenocarcinoma, 25% metastatic adenocarcinoma from the upper

gastrointestinal tract, and 40% metastatic adenocarcinoma of non-gastrointestinal origin and 20% metastatic carcinoma. Our results revealed that the sensitivity, specificity, positive predictive value, negative predictive value and accuracy of CDX2 were 80%, 72.5%, 59.3%, 87.9% and 75% respectively for the diagnosis of colorectal carcinoma metastasis from other metastatic carcinomas. These findings were in agreement with Dabir et al. (2018) who reported that CDX2 is used for identifying colorectal metastatic carcinomas, but many cases of other metastatic carcinomas showed positivity with this immunomarker. The calculated sensitivity and specificity of CDX2 in their study for colorectal metastatic carcinoma diagnosis were 93% and 67% respectively.

Bayrak et al. (2012) demonstrated that the sensitivity, specificity, positive predictive value, and negative predictive value of CDX2 expression and CK7-/CK20+ immunophenotyping to distinguish colonic carcinoma from gastric and pancreatic carcinomas. CK7/CK20 immunohistochemistry was more specific than CDX2 expression in the differentiation of colorectal adenocarcinoma from pancreatic and gastric adenocarcinomas with specificity 96.7%. They stated that CDX2 expression had a higher sensitivity and higher negative predictive value than the cytokeratin immunophenotype. The high sensitivity of CDX2 makes it useful marker for determining primary of metastatic colorectal carcinoma, but its specificity may be of concern.

Saad et al. (2013) noticed that CDX2 is a helpful immunomarker for colorectal metastatic adenocarcinoma. However, expression of CDX2 may be present in other tumors, especially tumors with intestinal differentiation, regardless of their site. They recommended that CDX2 could not be used alone for the detection of the primary origin of metastatic colorectal adenocarcinomas, should be used as a part of the immunohistochemical panel.

HNF4 α is one of the nuclear receptor family of transcription factors. Its location on chromosome 20 is regulated by two promoters (P1, P2). HNF4 α expression has been detected in liver, kidney and intestine. It has a role in liver and intestinal development (Davison et al.,

2017). Besides, its biological action in colorectal tissue, HNF4 α showed expression in CRC. HNF4 α is detected as the main transcription factor for colonic functions in both physiological and pathological conditions (Yao et al., 2016).

HNF4 α is the new marker that showed high specificity in metastatic colorectal adenocarcinoma. In the current study, HNF4 α was expressed in 18 of 20 (90%) colorectal adenocarcinoma, and 20% metastatic adenocarcinoma from upper gastrointestinal tract, 2 of 10 (20%) metastatic adenocarcinoma of non-gastrointestinal origin and 10% metastatic carcinoma. The calculated sensitivity, specificity, positive predictive value, negative predictive value and accuracy of HNF4 α were 90%, 82.5%, 72%, 94.3% and 85% respectively for diagnosis of colorectal carcinoma metastasis from other metastatic carcinomas. Our results suggested that HNF4 α is a novel marker that showed higher sensitivity and specificity in colorectal carcinoma metastasis than CDX2.

Kriegsmann et al. (2018) stated that HNF4 α may be useful in the differentiation of pulmonary adenocarcinoma and metastatic colorectal carcinoma. They found that HNF4 α exhibit positive expression in 77 of 80 cases (96%) of colorectal adenocarcinoma. However, they noticed this marker was not shown an improvement over the routinely used CDX2 and CK20.

In contrast to the current results, Tanaka et al. (2006) and Koyama et al. (2011) noted that HNF4 α is not specific to gastrointestinal adenocarcinomas, as positive immunoreactivity in some cases of renal cell carcinomas, HCC, and ovarian neoplasms may be present. Tanaka et al., 2006 claimed that HNF4 α immunomarker showed difference in neoplastic immunoreactivity for two isoforms. For instance, all 10 examined cases of renal cell carcinomas were positive for P1 and negative for P2. While gastric and colonic carcinomas were positive for P2 and less positive for P1. The difference may be due to variation of samples, difference in evaluation techniques, different isoforms, and included tumors variables.

In our study, HNF4 α was expressed in 4 of 20 (20%) metastatic adenocarcinoma from the

upper gastrointestinal tract. Moore et al. (2016) and Collepriest et al. (2017) explained HNF4 α expression in some upper gastrointestinal tract carcinoma cases, as HNF4 α showed positivity in gastric carcinoma cases with intestinal metaplasia. HNF4 α was also expressed in esophageal goblet cell metaplasia (Barrett esophagus). They found that Overexpression of HNF4 α in esophageal epithelium is suggestive of columnar phenotype of esophageal carcinoma.

We concluded that using HNF4 α as a supplementary marker with CDX2 to detect metastatic colorectal adenocarcinoma is helpful. HNF4 α expression is a highly specific and sensitive marker of colorectal origin. HNF4 α may be a useful immunomarker for detection of intestinal adenocarcinomas.

Conflict of interest

The authors claim no conflict of interest.

References

- Babeu JP, Boudreau F. (2014). Hepatocyte nuclear factor 4-alpha involvement in liver and intestinal inflammatory networks. *World J Gastroenterol*, 20: 22-30 .
- Bayrak, R., Haltas, H. & Yenidunya, S. (2012). The value of CDX2 and cytokeratins 7 and 20 expression in differentiating colorectal adenocarcinomas from extraintestinal gastrointestinal adenocarcinomas: cytokeratin 7-/20+ phenotype is more specific than CDX2 antibody. *Diagn Pathol*, 7, 9.
- Borrisholt M, Nielsen S and Vyberg M. (2013). Demonstration of CDX2 is highly antibody dependant. *Appl Immunohistochem Mol Morphol*, 21:64–72
- Chellappa K, Deol P, Evans JR, Vuong LM, Chen G, Briançon N, Bolotin E, Lytle C, Nair MG, Sladek FM. (2016). Opposing roles of nuclear receptor HNF4 α isoforms in colitis and colitis-associated colon cancer. *Elife*, 5.
- Collepriest BJ, Burke ZD, Griffiths LP, Chen Y, Yu WY, Jover R, Bock M, Biddlestone L, Quinlan JM, Ward SG, Mark Farrant J, Slack JM, Tosh D. (2017). HNF4 α is a key gene that can generate columnar metaplasia in oesophageal epithelium. *Differentiation*, 93:39-49.
- Dabir PD, Svanholm H, Christiansen JJ. (2018). SATB2 is a supplementary immunohistochemical marker to CDX2 in the diagnosis of colorectal carcinoma metastasis in an unknown primary. *APMIS*, 126: 494–500.
- Davison JM, Lickwar CR, Song L, Breton G, Crawford GE, Rawls JF. (2017). Microbiota regulate intestinal epithelial gene expression by suppressing the transcription factor hepatocyte nuclear factor 4 alpha. *Genome Res.*, 27; 1195–1206.
- Garrison WD, Battle MA, Yang C, Kaestner KH, Sladek FM, Duncan SA. (2006). Hepatocyte nuclear factor 4alpha is essential for embryonic development of the mouse colon. *Gastroenterology*, 130: 1207-1220 .
- Koyama T, Sekine S, Taniguchi H, Tsuda H, Ikegami M, Hano H, Kushima R. (2011). Hepatocyte nuclear factor 4A expression discriminates gastric involvement by metastatic breast carcinomas from primary gastric adenocarcinomas. *Hum Pathol.*, 42:1777-1784.
- Kriegsmann M, Harms A, Longuespee R. (2018). Role of conventional immunomarkers, HNF4 α and SATB2, in the differential diagnosis of pulmonary and colorectal adenocarcinomas. *John Wiley & Sons Ltd.*, 72:997-1006.
- Li MK, Folpe AL (2004). CDX-2, a new marker for adenocarcinoma of gastrointestinal origin. *Adv AnatPathol*, 11: 101-105.
- Lu H. (2016). Crosstalk of HNF4 α with extracellular and intracellular signaling pathways in the regulation of hepatic metabolism of drugs and lipids. *Acta Pharm Sin B*, 6: 393-408 .
- Moore BD, Khurana SS, Huh WJ, Mills JC. (2016). Hepatocyte nuclear factor 4 α is required for cell differentiation and homeostasis in the adult mouse gastric epithelium. *Am J Physiol Gastrointest Liver Physiol.*, 311:G267-G275.
- Ortiz-Rey JA, Alvarez C, San Miguel P, Iglesias B, Antón I (2005). Expression of CDX2, cytokeratins 7 and 20 in sinonasal intestinal type adenocarcinoma. *Appl Immunohistochem Mol Morphol.*, 13: 142-146.
- Park SY, Kim BH, Kim JH, Lee S, Kang GH (2007). Panels of immunohistochemical markers help determine primary sites of metastatic adenocarcinoma. *Arch Pathol Lab Med.*, 131: 1561-1567.
- Saad RS, Ghorab Z, Khalifa MA, Xu M. (2011). CDX2 as a marker for intestinal differentiation: Its utility and limitations. *World J Gastrointest Surg*, 3(11): 159-166.
- Saandi, T., Baraille, F., Derbal-Wolfrom, L., Cattin, A. L., Benahmed, F. (2013). Regulation of the tumor suppressor homeogene Cdx2 by HNF4 α in intestinal cancer. *Oncogene*, 32(32),3782-3788.
- Silberg DG, Swain GP, Suh ER, Traber PG (2000). Cdx1 and Cdx2 expression during intestinal development. *Gastroenterology.*, 119: 961-971.

Tanaka T, Jiang S, Hotta H (2006). Dysregulated expression of P1 and P2 promoter-driven hepatocyte nuclear factor-4alpha in the pathogenesis of human cancer. *J Pathol*, 208: 662–72.

Varadhachary GR (2007). Carcinoma of unknown primary origin. *Gastrointest Cancer Res.*, 1: 229-35.

Yao HS, Wang J, Zhang XP (2016). Hepatocyte nuclear factor 4alpha suppresses the aggravation of colon carcinoma. *Mol. Carcinog.*, 55; 458-472.

Egyptian Association for Cancer Research (EACR)

<http://eacr.tanta.edu.eg/>

EACR is an NGO society that was declared by the Ministry of Social Solidarity (Egypt) No. 1938 in 19/11/2014 based on the initiative of Prof. Mohamed Labib Salem, the current Chairman of EACR. EACR aims primarily to assist researchers, in particular young researchers in the field of cancer research through workshops, seminars and conferences. Its first international annual conference entitled "Anti-Cancer Drug Discovery" was successfully organized in April 2019 (<http://acdd.tanta.edu.eg>). Additionally, EACR aims to raise the awareness of the society about the importance of scientific research in the field of cancer research in prediction, early diagnosis and treatment of cancer. EACR is also keen to outreach the scientific community with periodicals and news on cancer research including peer-reviewed scientific journals for the publication of cutting-edge research. The official scientific journal of EACR is "International Journal of Cancer and biomedical Research (IJCBR: <https://jcbjournals.ekb.eg>)" was successfully issued in 2017 and has been sponsored by the Egyptian Knowledge Bank (EKB: www.ekb.eg).

EACR Chairman,

Prof. Mohamed Labib Salem, PhD

Professor of Immunology

Faculty of Science, Tanta University, Egypt

International Journal of Cancer & Biomedical Research
(IJCBR) Online ISSN 2682-2628

Editor-in-Chief

Mohamed Labib Salem, PhD
Tanta University, Egypt

Managing Editor

Nehal Elmashad, MD
Tanta University, Egypt
Nabil Mohy Eldin, PhD
Kafrelsheikh University, Egypt
Doaa Al-Ghareeb, PhD
Alexandria University, Egypt
Abdel-Aziz Zidan, PhD
Damanhour University, Egypt
Wesam Meshrif, PhD
Tanta University, Egypt
Rasha Eraky, MD
Tanta University, Egypt

Associate Editor

Hesham Tawfik
Tanta University, Egypt
Mostafa El-Sheekh
Tanta University, Egypt
Yousry Albolkin, PhD
Tanta University, Egypt
Gamal Badr
Assuit University, Egypt
Elsayed Salim
Tanta University, Egypt
Essam Elshiekh
Tanta Cancer Center, Egypt

Editorial Board

Alberto Montero
Taussig Cancer Center,
Cleveland, USA
Marcela Diaz
Cleveland Clinic Foundation, USA
Yi Zhang
Zhengzhou University, China
Shengdian Wang
Chinese Academy of Sciences,
China
Faris Alenzi
Prince Sattam bin Abdulaziz
University, KSA
Mark Robunstein
Medical University of South
Carolina, USA
Mamdooh Ghoneum, DSC
Charles Drew University of
Medicine & Science, USA

Natarajan Muthusamy, DVM
The Ohio State University, USA

Hideki Kasuya MD, PhD,
FACS
Nagoya University, Japan

Sherif El-Khamisy, MD
Sheffield University, UK

Mohamed Abou-El-Enein,
MD
Charité Universitätsmedizin
Berlin, Germany

Alaa Eldin Almostafa, MD
McGill University, Canada

Amr Amin
United Arab Emirates
University, UAE

AbdelRahman Zekri
National Cancer Institute, Egypt

Mohamed Attia, MD
Tanta University, Egypt

Mohamed Elshanshory, MD
Tanta University, Egypt

Hussein Khamis
Alexandria University, Egypt

Magdy Mahfouz
Kafr Elsheikh University, Egypt

Ehab Elbedewey
Tanta University, Egypt

Abeer Badr
Cairo University, Egypt

Nadia Hamdy, PharmD
Ain Shams University, Egypt

Ibrahim El-Sayed
Menoufia University, Egypt

Tarek Aboul-Fadl, PharmD
Assiut University, Egypt

Mohamed Nouredin
Banaha University, Egypt

Haiam Abou Elela
National Institute of
Oceanography and Fisheries,
Egypt

Sameh Ali, MD
Nationa Liver Institute, Egypt

Maha EL-Demellawi
City for Scientific Research &
Technology Applications, Egypt

Desouky A Abd-El-Haleem
City for Scientific Research &
Technology Applications, Egypt

Ashraf Tabll
National Research Center, Egypt

Wael Lotfy, MD
Alexandria University, Egypt

Olfat Gadallah, MD
Tanta University, Egypt

Nahla Shoukry
Suez University, Egypt

Medhat Eldenary
Tanta University, Egypt

Nagla Sarhan, MD
Tanta University, Egypt

Naglaa Fathy, MD
Zagazik University, Egypt

Azza Hasan Mohamed
Menoufia University, Egypt

Nanees Gamal Eldin
Tanta University, Egypt

Mohamed Mansour, UK

Sabbah Hammoury
Alexandria Ayadi Almostaqbal
Oncology Hospital, Egypt

Nehal Aboufotouh
Zewail City for Science and
Technology, Cairo, Egypt

Amir Elkhani
Galaxo, San Francisco, USA

Rabab Khairat
National Research Center,
Giza, Egypt

Ahmed Alzohairy
Zagazi University, Egypt

Wgady Khalil
National Research Center, Egypt

Sayed Bakry
Alazhar University, Egypt

Mohamed Ghanem, MD
Kafr Elshikh University, Egypt

Mohamed Salama, MD
Mansoura University, Egypt

Mona Marie, MD
Alexandria University, Egypt

For more information, contact

Hamdi Kandil
Tanta University, Egypt
Email: ljcb100@gmail.com