

Online ISSN: 2682-2628

Print ISSN: 2682-261X

IJC CBR

INTERNATIONAL JOURNAL OF CANCER AND BIOMEDICAL RESEARCH

<https://jcbr.journals.ekb.eg>

Editor-in-chief

Prof. Mohamed Labib Salem, PhD

**Measles vaccines may provide partial
protection against COVID-19**

Mahmoud E. Saad, Rokaya A. Elsalamony



PUBLISHED BY

EACR EGYPTIAN ASSOCIATION
FOR CANCER RESEARCH

Since 2014

**International Journal of Cancer & Biomedical Research
(IJCBR) <https://jcbr.journals.ekb.eg>**

IJCBR is an Int. journal published by the Egyptian Society of Cancer Research (EACR, established in 2014, <http://eacr.tanta.edu.eg>) and sponsored by the Egyptian Knowledge Bank (EKB: www.ekb.eg).

IJCBR has been approved by the Supreme Council of Universities, Egypt with score 7 (<http://egjournal.scu.eg>). The journal is cited by google scholar and registered by Publons (<https://publons.com>). The journal has recently been evaluated in 2020 by Nature Springer with a good standing.

Scope of IJCBR

- Drug discovery from natural and synthetic resources
- BioMedical applications of nanotechnology
- Sem cell biology and its application
- Basic and applied biotechnology
- Inflammation and autoimmune diseases
- In silico models and bioinformatics
- In vitro and In vivo preclinical animal models
- Cellular and molecular cancer biology
- Cancer Immunology and Immunotherapy
- New methods for prediction, early detection, diagnosis prognosis and treatment of diseases.
- Immunology in health and diseases
- Anti-microbial defense mechanisms
- Cellular and molecular physiology and pathology of diseases

IJCBR Editor,
Prof. Mohamed Labib Salem, PhD
Professor of Immunology
Faculty of Science, Tanta University, Egypt

Measles vaccines may provide partial protection against COVID-19

Mahmoud E. Saad¹, Rokaya A. Elsalamony²

¹Chemistry Department, Faculty of Science, Menoufia University, Menoufia, Egypt

²Biochemistry Department, Faculty of Science, Menoufia University, Menoufia, Egypt

ABSTRACT

In December 2019 a new coronavirus COVID-19 was identified in China then spread all over the world. WHO defined China and Italy as the epicenters for COVID-19. Insufficient vaccine coverage has been identified as a key causative factor in the most epidemic outbreaks. Vaccines generally raise specific immune responses to a targeted pathogen, but measles vaccines have recently proved the increased ability of the immune system to fight off pathogens other than measles. COVID-19 is proven to have similarities with measles. Such similarities may cause cross-reactivity between measles vaccines and COVID-19. For instance, comparing China and Italy for COVID-19 case and the death rates from late 2019 until Mars 25, 2020, Italy showed higher ratio of COVID-19 cases/population and a higher death rate than China. In contrast, Italy showed lower measles vaccination coverage than China. In this review, we hypothesized that the bystander immunity induced by measles vaccines may provide partial protection against COVID-19, decreasing the virus's ability to cause fatal symptoms and controlling the infection leading to full recovery. Accordingly, we suggest multi-center clinical trials to evaluate the possibility of induced partial protection by measles-containing vaccines against COVID-19.

Editor-in-Chief: Prof. M.L. Salem, PhD - Article DOI: 10.21608/IJCBR.2020.26765.1024

ARTICLE INFO



Article history

Received: March 28 2020

Revised: March 29 2020

Accepted: April 2, 2020

Correspondence to:

Dr. Mahmoud E. Saad
Chemistry Department,
Menoufia University, Egypt
Tel : +201021638998

E-Mail:

mahmoudsaadhegy@gmail.com

INTRODUCTION

In December 2019, patients who suffered from cough, fever, and dyspnea with acute respiratory distress syndrome (ARDS) due to an unknown viral infection were reported in Wuhan, China. Virus genome sequencing of five patients with pneumonia revealed the presence of a previously unknown coronavirus (β -CoV) strain in all of them [1,2].

Coronaviruses are enveloped viruses with a positive-sense single-stranded RNA genome (26–32 kb), which replicate in the cytoplasm of infected cells. There are four coronavirus genera (α , β , γ , δ) with human coronaviruses (HCoVs) detected in the α -coronavirus (HCoV-229E and NL63) and β coronavirus (MERS-CoV, SARS-CoV, HCoV-OC43 and HCoV-HKU1) genera [3].

The isolated novel β -CoV shows 88% similarity to the sequence of two bat-derived severe acute respiratory syndromes (SARS)-like coronaviruses, bat-SL-CoVZC45 and bat-SL-

CoVZXC21, and about 50% similarity to the sequence of MERS-CoV *4+. The novel β -CoV was then named "SARS-CoV-2" by the International Virus Classification Commission [5].

Viral structure

Viral particles are composed of four major structural proteins: the nucleoprotein (N), the small envelope protein (E), the membrane protein (M), and the large spike protein (S). The entry of coronaviruses into cells is mediated by the transmembrane spike glycoprotein S, which forms a trimer carrying receptor-binding and membrane fusion functions. The S protein (SP) also contains the principal antigenic determinants and is the target of neutralizing antibodies [6].

Coronaviruses Spike protein (SP)

CoVs use their spike (S) proteins for host recognition and subsequent membrane fusion to introduce their viral genomes into the host for replication. Preventing CoV infection by

blocking S-protein binding to host receptors : angiotensin-converting enzyme 2 (ACE2) for SARS-CoV, dipeptidyl peptidase 4 (DPP4) for MERS-CoV and Glucose Regulated Protein 78 (GRP78) for SARS-CoV-2[7,8], therefore, represents the first line of defense[7].

CoV S proteins consist of 2 functional units, the S1 and S2 subunits, which are responsible for cell attachment and membrane fusion, respectively [8,9]. Mutations in the receptor-binding motifs (RBMs) or cleavage sites of CoV S proteins can lead to zoonotic spillover and alteration of cell/tissue tropism, as exemplified by SARS and MERS [10,11].

The coronavirus SP is a class I viral fusion protein and considered the major viral regulator in host cell entry [12]. Viral fusion proteins have grouped into three different classes according to their structure and biochemical activation processes, where class I proteins are characterized by predominant α - helical secondary structures and a trimeric organization of their pre-fusion and post-fusion states [13,14].

One interesting aspect of the class I fusion proteins is the differences in activation of their fusion mechanisms, despite their conserved structures [15,16]. However, all fusion proteins in this class undergo major structural changes that allow the viral fusion peptide to contact and anchor into the target cell membrane, and the formation of the “trimer of hairpins” structure followed by the fusion of the outer membranes (hemifusion) and the formation of the fusion pore. To successfully induce fusion, proteolytic activation of the viral protein subunits is often necessary, and this can vary significantly between different fusion proteins [16].

Similarities between Coronaviruses and Paramyxoviruses

Paramyxovirus is any virus belonging to the family Paramyxoviridae. Paramyxoviruses have enveloped virions (virus particles). The paramyxovirus genome is made up of a single strand of negative-sense non segmented RNA (ribonucleic acid). An endogenous RNA polymerase is present as well as is necessary for the transcription of the negative-sense strand

into a positive-sense strand, thereby enables proteins to be encoded from the RNA. The lipoprotein envelope contains two glycoprotein spikes, including designated hemagglutinin-neuraminidase (HN) and fusion factor (F).

Paramyxoviridae has two subfamilies, *Paramyxovirinae* and *Pneumovirinae*, each of which contains multiple genera. Examples of *Paramyxovirinae* genera include *Rubula* virus, which is composed of several species of human parainfluenza viruses and the mumps viruses; *Avulavirus*, species Newcastle contains virus (of contains the poultry) and Morbilli virus, which disease which the agents that cause measles in humans. Species of *Pneumovirus*, which are responsible for the serious respiratory syncytial virus disease in human infants, are classified in the subfamily *Pneumovirinae* [17].

Coronavirus SP shares a common core with *paramyxovirus* F proteins [18], implicating mechanistic similarities and an evolutionary connection between these viral fusion proteins. The accessibility of the highly conserved fusion peptide at the periphery of the trimer indicates potential vaccine strategies to elicit broadly neutralizing antibodies against coronaviruses. Comparison with crystal structures of human coronavirus S domains allows rationalization of the molecular basis for species specificity based on the use of spatially contiguous but distinct domains [19]. Despite weak sequence conservation, the structure demonstrates structural similarity to paramyxovirus F proteins, thereby reinforcing the relatedness of their fusion mechanisms and their evolutionary connection, which provide a structural framework to rationalize the mode of antibodies neutralization targeting the conserved fusion machinery [19].

Cross-reactivity and cross-protection

Cross-reactivity is the reaction between an antibody and an antigen that differs from the immunogen. It is also referred to as cross-immunity or cross-protective immunity. Cross-protection is a phenomenon in which infections with a mild virus or viroid strain protects from disease resulting from a subsequent encounter with a severe strain of the same virus or strain [20].

Although cross-reactivity does not necessarily infer cross-protection, a few examples of cross-reactivity have been confirmed in humans, one of which involves influenza virus-specific CD8+ T cell and hepatitis C virus antigens [21].

Studies proved that humoral immunity elicited by huN1 (human H1N1 infection) can partially protect against H5N1 infection in a mammalian host, suggesting that a portion of the human population could have some degree of resistance to H5N1 influenza. This possibility could be induced or enhanced through immunization with seasonal influenza vaccines, which was confirmed by studies in humans [22].

Measles containing vaccines (MCV)

Measles virus (MV) belongs to the genus Morbillivirus of the family Paramyxoviridae. Although these viruses are distinct agents, they share certain antigens [23]. MV vaccine (MCV) is a live-attenuated negative-stranded RNA virus was proven to be one of the safest and most effective human vaccines. This vaccine is produced on a large scale in many countries and distributed at low cost through the Extended Program on Immunization (EPI), induces life-long immunity to measles after one or two injections [24].

MCVs were first licensed in 1963. Since the 1980s, several live attenuated measles vaccines were developed, either as a monovalent vaccine or in combination with rubella, mumps, or varicella vaccines, or some combinations of these. When using the combined measles–rubella (MR) vaccine, measles–mumps–rubella (MMR) vaccine, or measles–mumps–rubella–varicella (MMRV) vaccine, the protective immune response to each individual vaccine antigen is largely unchanged [24].

Over the last decade, outbreaks of vaccine-preventable diseases have been reported in developed countries around the world. Measles outbreaks have been ongoing in the European Union, with most cases concentrated in Romania and Italy. Measles has been identified as a powerful indicator of the status of vaccination programs in a region, as outbreaks have been reported to quickly emerge as a result of underlying problems in the immunization routine.

Insufficient vaccine coverage has been identified as a key causative factor in most measles outbreaks. Italy accounts for over 30% of measles cases reported since 2017 in the European Union [24]. In Italy mean measles vaccination coverage from 1980 until 2019 is 84.5%. The majority of reported measles cases in 2016 were of unvaccinated people. One-third of the cases were among children aged 1–4 years [25].

China is making significant progress in battling measles disease, according to a December report from the Centers for Disease Control and Prevention. From 2013–2018, measles vaccine coverage in the country was estimated at 99%. Measles incidence declined from 31 per million in 2015 to 2.8 in 2018; only one measles-associated death has been reported during 2018–June 2019 [26].

Reduction effect of Measles vaccine on all-cause mortality

Vaccines generally raise immune responses specific to a targeted pathogen, such as antibodies that bind and neutralize one type of virus but not others. But MVC vaccine may also increase the ability of the immune system to fight off pathogens other than measles. According to a recent study, receipt of MCV standard titer was associated with a reduction in all-cause mortality (relative risks 0.74:0.51 to 1.07%) from four clinical trials and (0.51: 0.42 to 0.63%) from 18 observational studies. A study suggests that receipt of MCV reduces overall mortality through their effects on the diseases that they prevent [27].

COVID-19 and measles vaccines

WHO defined China as the first epicenter for COVID-19, then Italy became the second with an even higher death rate. Comparing the number of COVID-19 cases and the associated death rates in China and Italy from late 2019 until March 25, 2020 [25, 28] showed that Italy has lower as significantly higher ratio of COVID-19 cases/population (57 in China versus 1,431 in Italy) and a higher death rate than China (2 in China versus 151 in Italy).

When comparing the measles vaccination coverage in the two countries, China (96.7%) had higher coverage than Italy (84%). According

to this profile, we suggest that the significant higher COVID-19 cases/population ratio and higher death rate in Italy as compared to China may be, at least in part, due to the lower measles vaccination coverage in Italy than china.

Perspective

From comparing statistical data for measles vaccination coverage, COVID-19 cases and deaths, we suggest that the MCV may provide partial protection against COVID-19. This vaccination induces immune system to fight the infection, to decrease the virus's ability to cause fatal symptoms and to control the infection leading to full recovery. Consistent with our suggestion, routine childhood vaccination, such as BCG, has also been suggested to provide bystander immunity to combat COVID-19 [29,30]. We suggest the following two mechanisms that explain the ability of MCV to cause partial protection against COVID-19. The first is that MCV may increase the ability of the immune system to fight off pathogens other than measles due to the generated bystander immunity that would enhance the overall immunity against the new coronavirus. The second is that COVID-19 is proven to have structure similarities with measles, which may cause cross-reactivity and immunity between measles vaccines and COVID-19, leading to partial protection against COVID-19 in vaccinated subjects.

Recommendations

We recommend designing multi-center clinical trials to evaluate the possibility of induction of partial protection by measles-containing vaccines against COVID-19.

Conflict of interest

Authors declare that they have no conflicts of interest.

References

1. Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*. 2020; 395: 497-506.
2. Zhong N, Zheng B, Li Y, PoonL, Xie Z, Chan K, et al. Epidemiology and cause of severe acute respiratory syndrome (SARS) in Guangdong,

- People's Republic of China. *The Lancet*, 2003, 362 (9393): 1353-1358
3. Wu F, Zhao S, Yu B et al. A new coronavirus associated with human respiratory disease in China. *Nature*. 2020; (published online Feb 3.).
4. Lu R, Zhao X, Li J et al. Genomic characterization and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding, *Lancet* (2020),10.1016/S0140-6736(20)30251-8
5. LiX,GengM,PengY,MengL,LuS. Molecular immune pathogenesis and diagnosis of COVID-19. *Journal of Pharmaceutical Analysis*. Available online 5 March 2020.
6. Burkard, C. et al. Coronavirus cell entry occurs through the endo-/lysosomal pathway in a proteolysis-dependent manner. *PLoS Pathog*. 2014, 10, e1004502).
7. de Wit E, van Doremalen N, Falzarano D et al. SARS and MERS: recent insights into emerging coronaviruses. *Nat. Rev. Microbiol.*, 2016, 14: 523-534.
8. Ibrahim IM, Abdelmalek DH, Elshahat ME, Elfiky AA. COVID-19 spike-host cell receptor GRP78 binding site prediction. *J Infect*. 2020, 10, pii:S0163-4453(20)30107-9.
9. Spiga O, Bernini A, Ciutti A, Chiellini S, Menciassi N, Finetti F, Causarano V, Anselmi F, Prischi F and Niccolai. Molecular modelling of S1 and S2 subunits of SARS coronavirus spike glycoprotein. *Biochem. Biophys. Res. Commun*. 2003, 310:78.
10. Millet JK and Whittaker GR. Host cell proteases: Critical determinants of coronavirus tropism and pathogenesis. *Virus Res*. 2015, 202: 120–134.
11. H. D. Klenk, W. Garten, Host cell proteases controlling virus pathogenicity. *Trends Microbiol*. 1994, 2: 39–43.
12. Bosch BJ, van der Zee R, Cornelis de Haan AM, Rottier PJM. The Coronavirus Spike Protein Is a Class I Virus Fusion Protein: Structural and Functional Characterization of the Fusion Core Complex. *J Virol*. 2003, 77(16): 8801-11.
13. Knipe DM and Howley PM (Eds.). *Harrison Principles of virus structure*. Fields Virology (6th ed.), Lippincott Williams & Wilkins, Philadelphia, PA, 2013: 52-86
14. White JM, Delos SE, Brecher M, Schornberg K. Structures and mechanisms of viral membrane fusion proteins: multiple variations on a common theme. *Crit. Rev. Biochem. Mol. Biol.*, 2008, 43: 189-219
15. Millet JK, Whittaker GR. Host cell proteases: critical determinants of coronavirus tropism and pathogenesis. *Virus Res.*, 2015, 202: 120-134

16. McLellan JS et al. Structure of RSV fusion glycoprotein trimer bound to a prefusion-specific neutralizing antibody. *Science* 2013, 340, 1113–1117.
17. Enders G. Paramyxoviruses. In: Baron S, editor. *Medical Microbiology*. 4th edition. Galveston (TX): University of Texas Medical Branch at Galveston; 1996. Chapter 59.
18. Yin HS, Wen X, Paterson RG, Lamb RA and Jardetzky TS. Structure of the parainfluenza virus 5 F protein in its metastable, prefusion conformation. *Nature*, 2006, 439: 38–44.
19. Walls AC, Tortorici AM, Bosch B, Frenz B, Rottier PJM, DiMaio F, Rey FA and Vesler D. Cryo-electron microscopy structure of a coronavirus spike glycoprotein trimer. *Nature* volume, 2016, 531: 114–117.
20. Ziebell H and Carr JP. *Advances in Virus research*, 2010, 76: 211-264
21. Kasproicz V, Ward SM, Turner A, Grammatikos A, Nolan BE, Lewis-Ximenez L, Sharp C, Woodruff J, Fleming VM, Sims S, Walker BD, Sewell AK, Lauer GM, Klenerman P. Defining the directionality and quality of influenza virus-specific CD8+ T cell cross-reactivity in individuals infected with hepatitis C virus. *J Clin Invest*. 2008,118(3):1143-53
22. Sandbulte MR1, Jimenez GS, Boon AC, Smith LR, Treanor JJ, Webby RJ. Cross-reactive neuraminidase antibodies afford partial protection against H5N1 in mice and are present in unexposed humans. *PLoS Med*. 2007 Feb;4(2):e59.
23. Rima BK1, Duprex WP. Morbilliviruses and human disease. *J Pathol*. 2006, 208(2):199-214.
24. Alessandro Siani. Measles outbreaks in Italy: A paradigm of the re-emergence of vaccine-preventable diseases in developed countries, *Preventive Medicine*. 2019, 121: 99-104.
25. WHO/UNICEF Joint Reporting Forms (JRFs) for disease incidence and WHO-UNICEF estimates of national immunization coverage (WEUNIC) data for coverage rates. https://www.who.int/immunization/monitoring_surveillance/routine/reporting/en.
26. Ma C, Rodewald L, Hao L, Su Q, Zhang Y. Progress Toward Measles Elimination — China, January 2013–June 2019, *CDC Weekly*, December 6, 2019, 68(48);1112– 1116
27. Higgins JPT, Soares-Weiser K, López-López JA, Kakourou A, Chaplin AK, Christensen H, Martin NK, Sterne JAC, Reingold AL. Association of BCG, DTP, and measles containing vaccines with childhood mortality: systematic review, *BMJ* 2016;355:i5170.
28. Coronavirus-Worldometer. <https://www.worldometers.info/coronavirus>. March 25, 2020.
29. Salman, S., Salem, M.L. (2020). The mystery Behind Childhood Sparing by COVID-19, *Int. J. Cancer and Biomedical Research*, 2020, 5(1):11-13.
30. Salman S. and Salem M.L.(2020), Routine childhood immunization may protect against COVID-19. Volume 140, July 2020, 109689

Egyptian Association for Cancer Research (EACR)

<http://eacr.tanta.edu.eg/>

EACR is an NGO society that was declared by the Ministry of Social Solidarity (Egypt) No. 1938 in 19/11/2014 based on the initiative of Prof. Mohamed Labib Salem, the current Chairman of EACR. EACR aims primarily to assist researchers, in particular young researchers in the field of cancer research through workshops, seminars and conferences. Its first international annual conference entitled "Anti-Cancer Drug Discovery" was successfully organized in April 2019 (<http://acdd.tanta.edu.eg>). Additionally, EACR aims to raise the awareness of the society about the importance of scientific research in the field of cancer research in prediction, early diagnosis and treatment of cancer. EACR is also keen to outreach the scientific community with periodicals and news on cancer research including peer-reviewed scientific journals for the publication of cutting-edge research. The official scientific journal of EACR is "International Journal of Cancer and biomedical Research (IJCBR: <https://jcbjournals.ekb.eg>) was successfully issued in 2017 and has been sponsored by the Egyptian Knowledge Bank (EKB: www.ekb.eg).

EACR Chairman,

Prof. Mohamed Labib Salem, PhD

Professor of Immunology

Faculty of Science, Tanta University, Egypt

International Journal of Cancer & Biomedical Research
(IJCBR) Online ISSN 2682-2628

Editor-in-Chief

Mohamed Labib Salem, PhD
Tanta University, Egypt

Managing Editor

Nehal Elmashad, MD
Tanta University, Egypt
Nabil Mohy Eldin, PhD
Kafrelsheikh University, Egypt
Doaa Al-Ghareeb, PhD
Alexandria University, Egypt
Abdel-Aziz Zidan, PhD
Damanhour University, Egypt
Wesam Meshrif, PhD
Tanta University, Egypt
Rasha Eraky, MD
Tanta University, Egypt

Associate Editor

Hesham Tawfik
Tanta University, Egypt
Mostafa El-Sheekh
Tanta University, Egypt
Yousry Albolkin, PhD
Tanta University, Egypt
Gamal Badr
Assuit University, Egypt
Elsayed Salim
Tanta University, Egypt
Essam Elshiekh
Tanta Cancer Center, Egypt

Editorial Board

Alberto Montero
Taussig Cancer Center,
Cleveland, USA
Marcela Diaz
Cleveland Clinic Foundation, USA
Yi Zhang
Zhengzhou University, China
Shengdian Wang
Chinese Academy of Sciences,
China
Faris Alenzi
Prince Sattam bin Abdulaziz
University, KSA
Mark Robunstein
Medical University of South
Carolina, USA
Mamdooh Ghoneum, DSC
Charles Drew University of
Medicine & Science, USA

Natarajan Muthusamy, DVM
The Ohio State University, USA

Hideki Kasuya MD, PhD,
FACS
Nagoya University, Japan

Sherif El-Khamisy, MD
Sheffield University, UK

Mohamed Abou-El-Enein,
MD
Charité Universitätsmedizin
Berlin, Germany

Alaa Eldin Almostafa, MD
McGill University, Canada

Amr Amin
United Arab Emirates
University, UAE

AbdelRahman Zekri
National Cancer Institute, Egypt

Mohamed Attia, MD
Tanta University, Egypt

Mohamed Elshanshory, MD
Tanta University, Egypt

Hussein Khamis
Alexandria University, Egypt

Magdy Mahfouz
Kafr Elsheikh University, Egypt

Ehab Elbedewey
Tanta University, Egypt

Abeer Badr
Cairo University, Egypt

Nadia Hamdy, PharmD
Ain Shams University, Egypt

Ibrahim El-Sayed
Menoufia University, Egypt

Tarek Aboul-Fadl, PharmD
Assiut University, Egypt

Mohamed Nouredin
Banaha University, Egypt

Haiam Abou Elela
National Institute of
Oceanography and Fisheries,
Egypt

Sameh Ali, MD
Nationa Liver Institute, Egypt

Maha EL-Demellawi
City for Scientific Research &
Technology Applications, Egypt

Desouky A Abd-El-Haleem
City for Scientific Research &
Technology Applications, Egypt

Ashraf Tabll
National Research Center, Egypt

Wael Lotfy, MD
Alexandria University, Egypt

Olfat Gadallah, MD
Tanta University, Egypt

Nahla Shoukry
Suez University, Egypt

Medhat Eldenary
Tanta University, Egypt

Nagla Sarhan, MD
Tanta University, Egypt

Naglaa Fathy, MD
Zagazik University, Egypt

Azza Hasan Mohamed
Menoufia University, Egypt

Nanees Gamal Eldin
Tanta University, Egypt

Mohamed Mansour, UK

Sabbah Hammoury
Alexandria Ayadi Almostaqbal
Oncology Hospital, Egypt

Nehal Aboufotouh
Zewail City for Science and
Technology, Cairo, Egypt

Amir Elkhani
Galaxo, San Francisco, USA

Rabab Khairat
National Research Center,
Giza, Egypt

Ahmed Alzohairy
Zagazi University, Egypt

Wgady Khalil
National Research Center, Egypt

Sayed Bakry
Alazhar University, Egypt

Mohamed Ghanem, MD
Kafr Elshikh University, Egypt

Mohamed Salama, MD
Mansoura University, Egypt

Mona Marie, MD
Alexandria University, Egypt

For more information, contact

Hamdi Kandil
Tanta University, Egypt
Email: ljcb100@gmail.com